



PROJECT 19 06 02 02

Development of research infrastructure and techniques at
IFIN-HH Tandem accelerators with
new experimental setups and analytical methods

PN Responsible: Gihan Velisa



CORE PROJECT: 19 06 02 02

Phase 6:
Designing and developing a new ion beam setup for
radiobiology studies (I & II)

Mihai Straticiuc, Mihaela Bacalum, Călin Mircea Rusu, Radu Andrei, Ion Burducea, Ioan Cenusă,
Constantin Cenusă, Irina Dinescu, Simona Dirleci, Alexandru Enciu, Decebal Iancu, Radu Vasilache, Mina
Raileanu, Mihai Radu

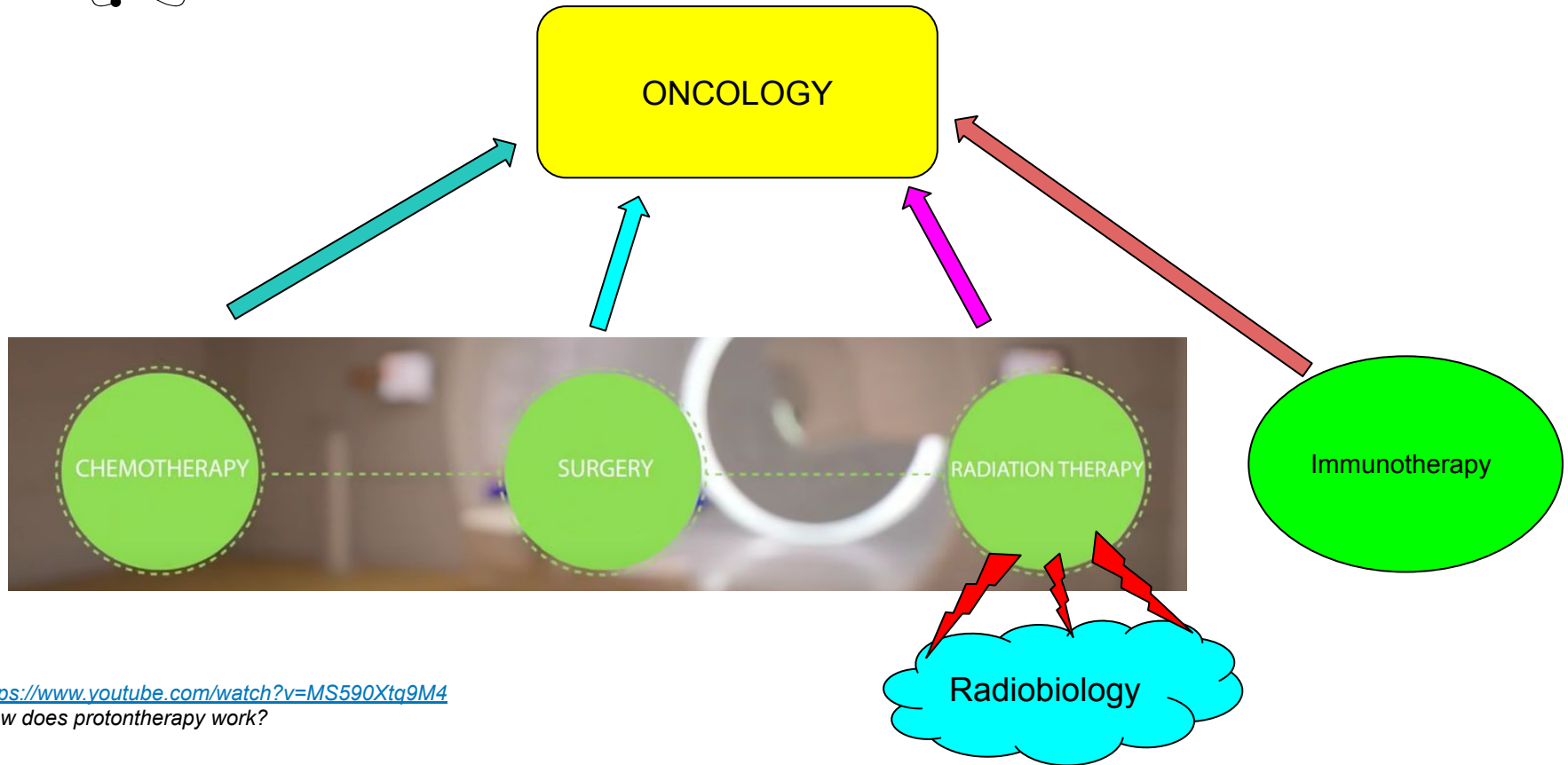


Project objectives:

1. Innovative technological developments at the Tandem accelerators, in order to maintain the capacity for analysis and irradiation at the highest level.
2. Technological developments of the experimental area, in order to meet the new needs of the users of the research infrastructure.
3. Development of new techniques and technologies that will find their applicability in the industrial, environmental and/or security area.
4. Prototype developments for international projects where IFIN-HH act as a member.



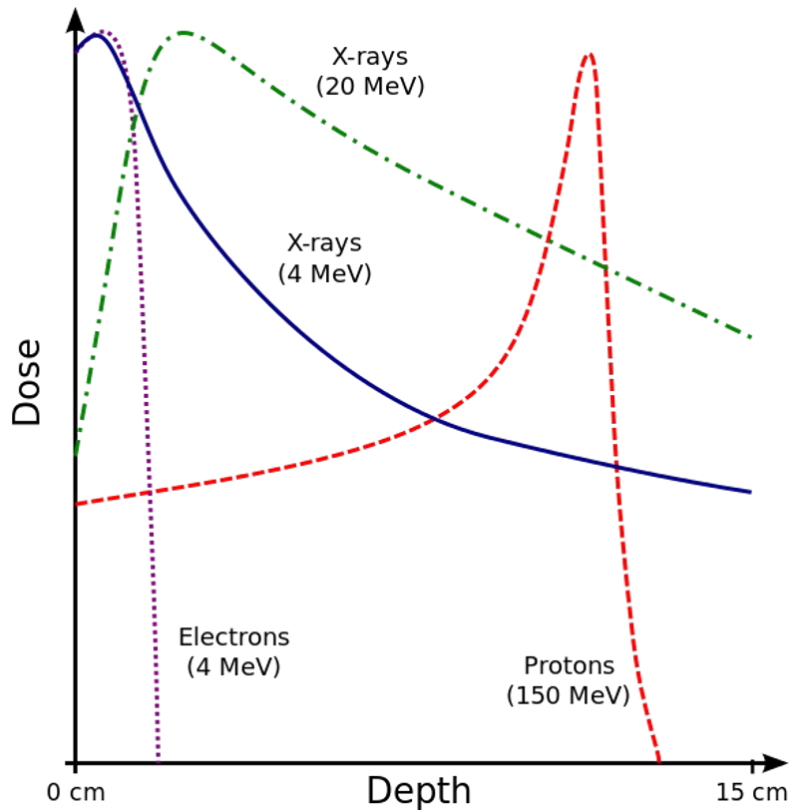
MOTIVATION



<https://www.youtube.com/watch?v=MS590Xtg9M4>
How does protontherapy work?



MOTIVATION: photons vs ions



Beer-Lambert (photons):

$$I = I_0 * e^{-\mu x}$$

Bethe-Bloch (ions):

$$-\frac{dE}{dx} = \frac{4\pi n z^2}{m_e v^2} \cdot \left(\frac{e^2}{4\pi\epsilon_0}\right)^2 \cdot \left[\ln\left(\frac{2m_e v^2}{I}\right)\right]$$

Radiological Use of Fast Protons

Robert R. Wilson

Published Online: Nov 1 1946 | <https://doi.org/10.1148/47.5.487>

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Tools Share



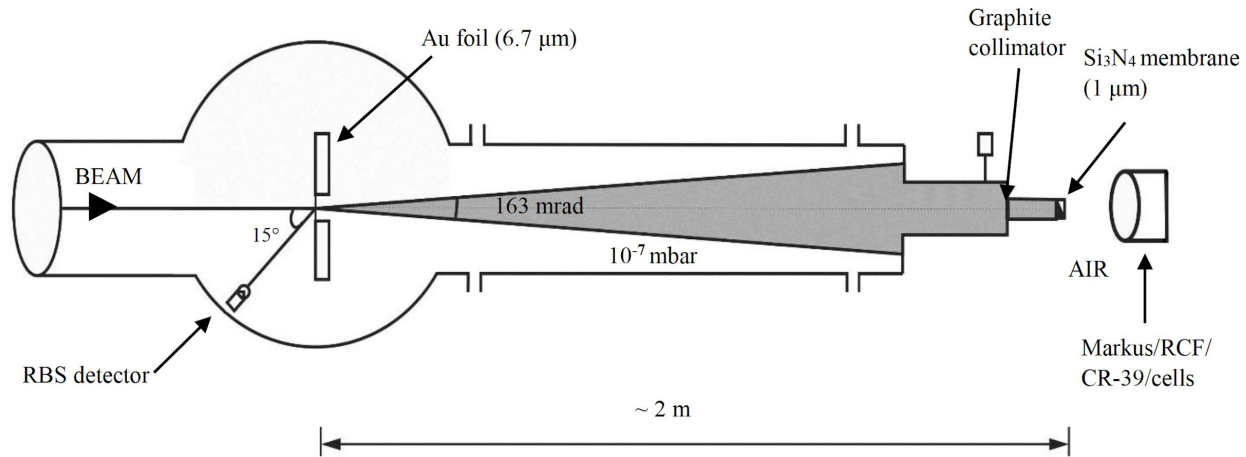
3 MV HVE Tandetron™ accelerator installed at IFIN-HH

I. Burducea et al., NIM B, 359, 12-19, 2015

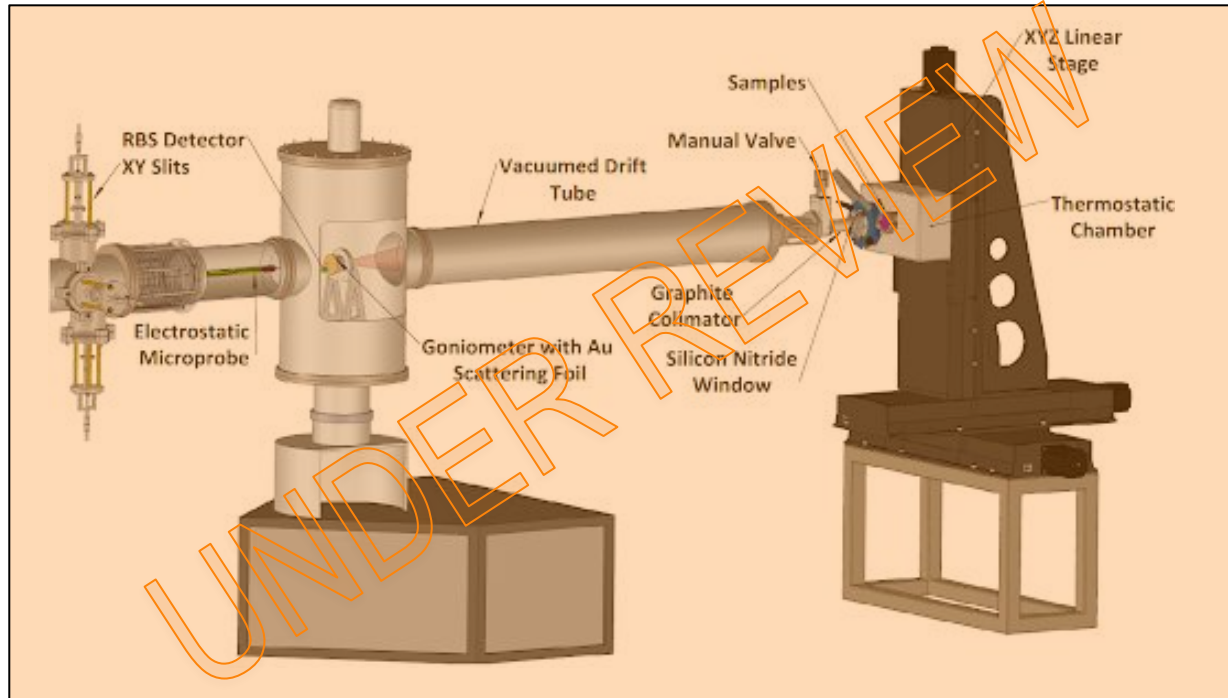


Photo credit: A. Socolov

2D Experimental Setup (v1)



3D Experimental Setup (v2)



Courtesy of A. Enciu

Experimental Setup (6th of December 2021)

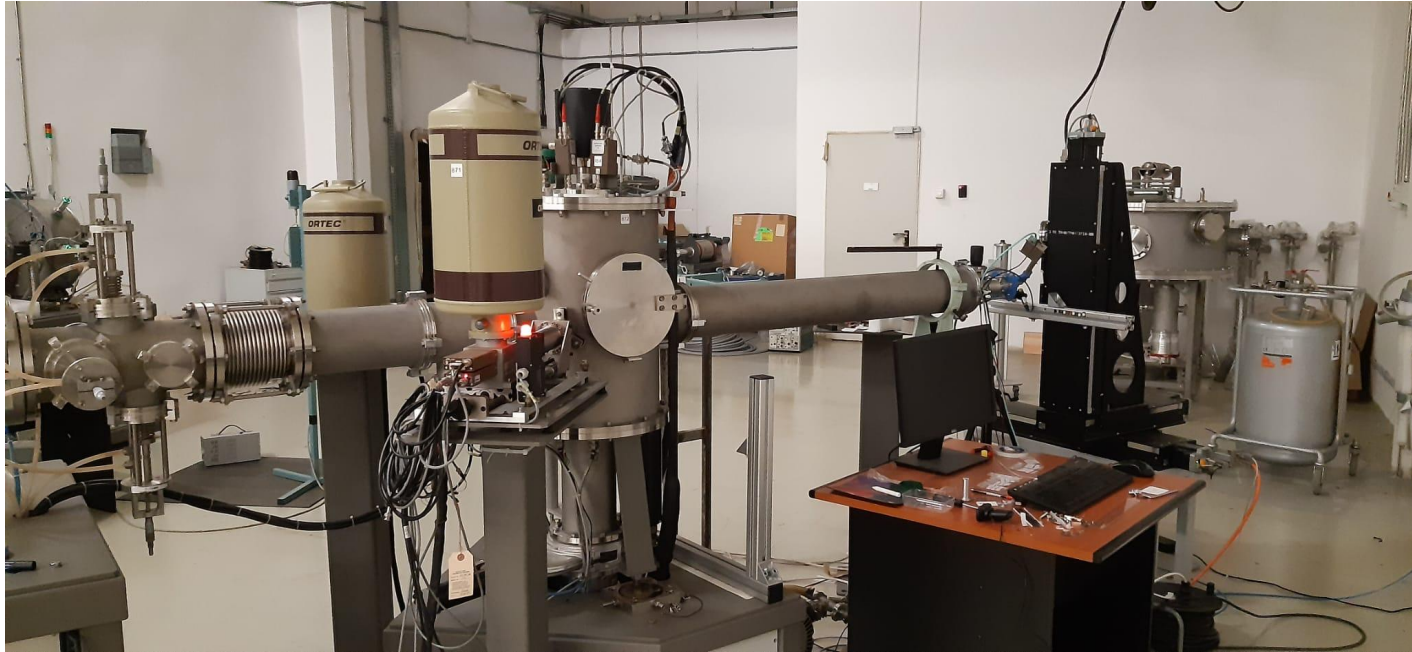
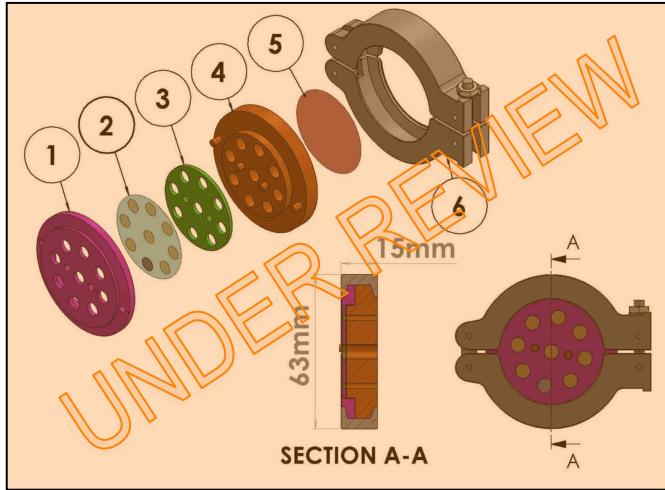


Photo credit: D. Iancu



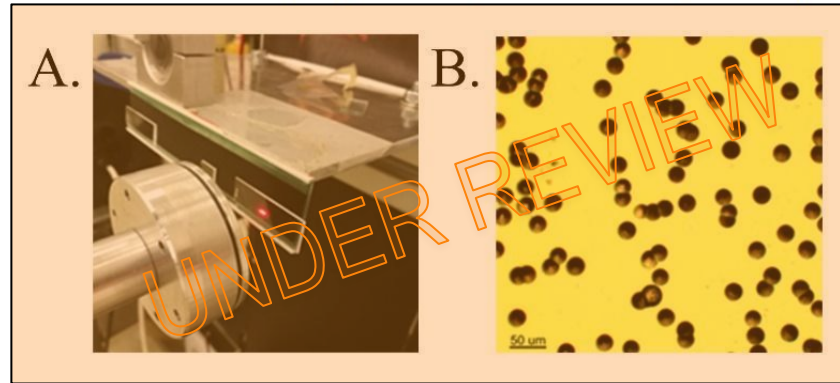
Cell culture "homemade" chamber &

1st dosimetry attempts with Tastrak® detectors



Scheme of a cell culture chamber (1. Aluminum front cover; 2. Mylar foil; 3. Silicone gasket; 4. Teflon chamber; 5. Breathable sealing tape sterile; 6. KF63 vacuum clamp)

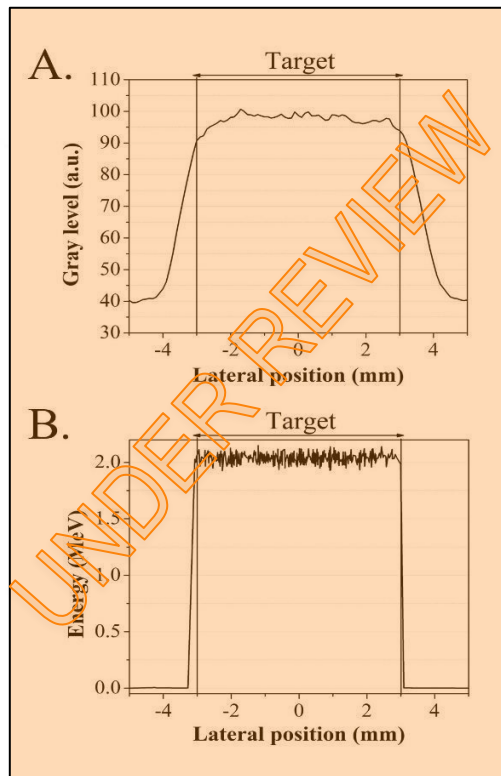
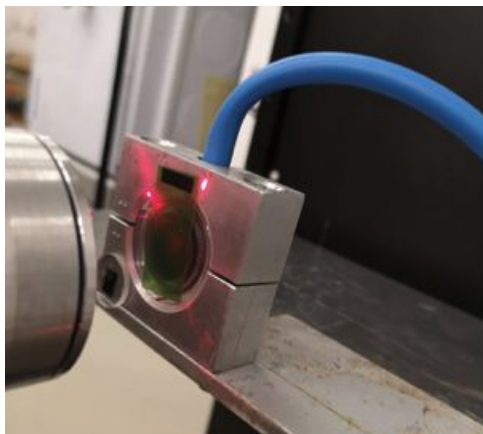
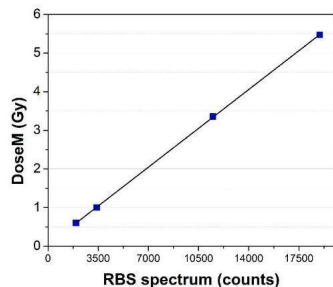
$$D [\text{Gy}] = 1.6 * 10^{-9} * \phi (\text{p/cm}^2) * dE/dx (\text{keV}/\mu\text{m})$$



A. Tastrak® detectors irradiation (1. Si_3N_4 window holder; 2. Tastrak® detector); B. photo taken under microscope after etching procedure was applied.

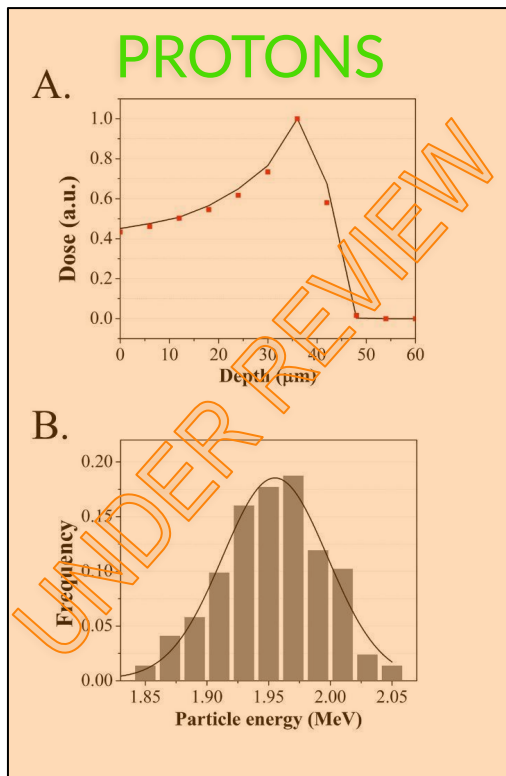


Beam dosimetry (Markus ionization chamber, RCF & Si particle detector)



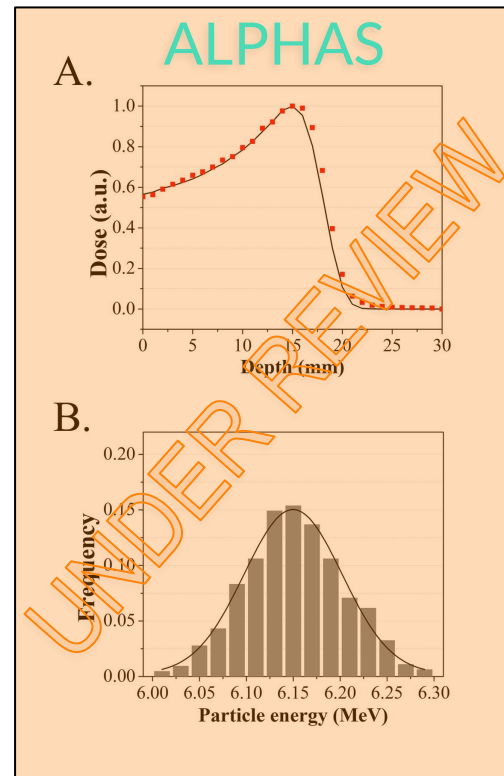
A. Lateral uniformity of the beam as recorded with radiochromic film expressed as gray level distribution along the spot diameter; B. Lateral energy distribution simulated with Geant4 expressed as the energy profile along the diameter at the air target interface in the same position as the film.

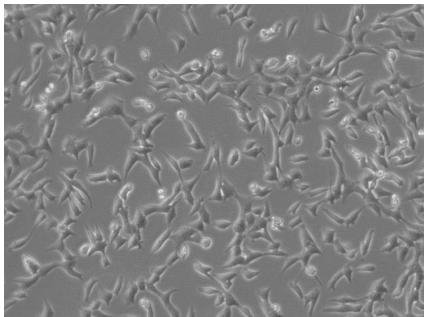
Depth dose profile measured with Markus vs Geant4 simulation



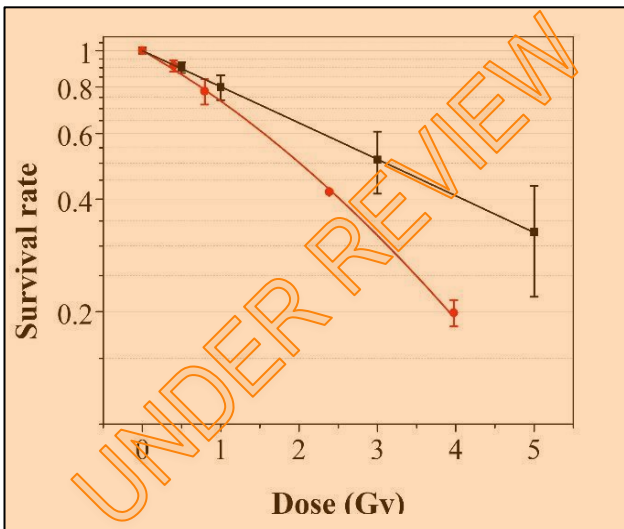
A. Experimental recording (red points) and Geant4 simulation (line plot) of protons/alphas Bragg peak in Mylar/air, $x = 0$ represents the air-mylar interface position during irradiation

B. energy distribution of the proton beam at the air-Mylar interface and Gauss fit (LEFT) and energy distribution of the alpha beam at the air-Mylar interface (RIGHT).





V79-4 cells 1d after seeding in EMEM with 10% FBS



Cell survival rate of V79 cells irradiated with doses between 0-5 Gy. The red line and dots represent the survival rate for protons. The black line and dots represent the survival rate for X-rays.

$$SF = \exp(-\alpha D - \beta D^2)$$

SF- survival fraction

D - dose (Gy)

α , β - linear & quadratic fit coefficients

Radiation type	$\alpha \pm \text{sd} (\text{Gy}^{-1})$	$\beta \pm \text{sd} (\text{Gy}^{-2})$	$D_{50}; (*D_{10}) (\text{Gy})$	RBE 50; (RBE 10)
Protons	0.27 ± 0.02	0.035 ± 0.007	2; (5)	1.5; (2)
X rays	0.22 ± 0.005	0.0005 ± 0.002	3; (10)	1.00

* D_{10} was estimated by fitting function.

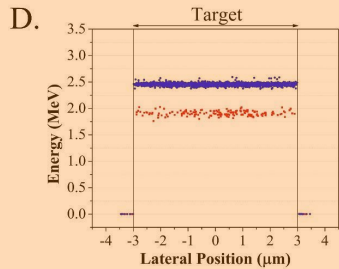
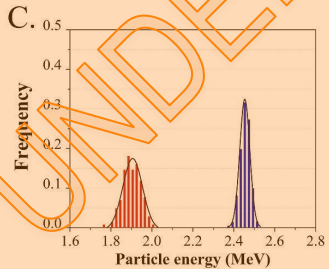
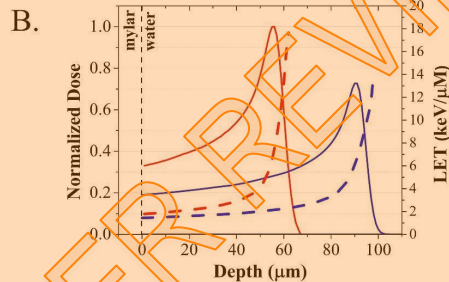
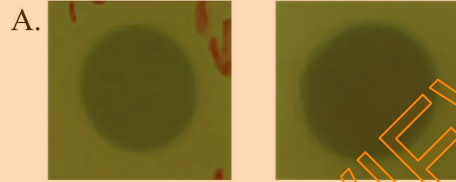


Conclusions

- a new cell culture irradiation setup for basic radiobiological investigations and preliminary experimental results obtained for the first time at IFIN-HH 3 MV TandatronTM are reported.
- the proton beam was extracted into the air through a Si_3N_4 membrane, and the dose was estimated by several methods.
- preliminary cell survival results show a similarity to data reported in the literature using other proton beam facilities for V79 cells.
- Geant4 simulations confirmed the experimental data of the Bragg peak for two ion species (protons and alpha) and provided promising predictions for switching to the FLASH regime and also the use of high ion beams with high LET (${}^7\text{Li}^{3+}$, ${}^{11}\text{B}^{5+}$ and ${}^{12}\text{C}^{6+}$) which could be supplied by IFIN-HH's 9 MV Van de Graaff tandem accelerator.

Ultra-High Dose Rate (FLASH) Radiotherapy: Silver Bullet or Fool's Gold?

Joseph D Wilson¹, Ester M Hammond¹, Geoff S Higgins¹, Kristoffer Petersson^{1 2}



A. RCF irradiated at 100 Gy/s (left) RCF irradiated at 250 Gy/s (right);

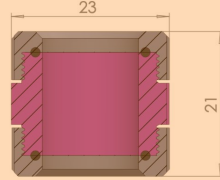
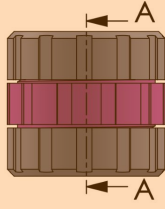
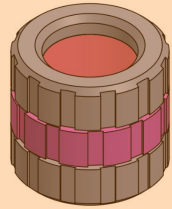
B. Geant4 simulation of dose (line plot) and LET (dashed plot) in water after protons propagation through different thin foils; red: low flux protons through the 6.7 μm Au foil, blue: high flux protons through the 2 μm Al foil;

C. Lateral energy distributions of the low and high flux proton beams at the Mylar-water interface simulated by Geant4 expressed as the energy profile along the target diameter;

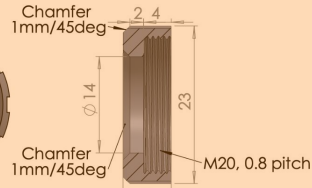
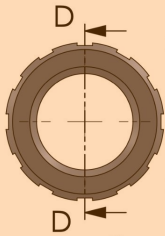
D. Energy distribution of the low and high flux proton beams at the mylar-water interface (low flux: mean energy = 1.9 MeV, FWHM = 104.02 keV; high flux: mean energy = 2.45 MeV, FWHM = 56.18 keV).



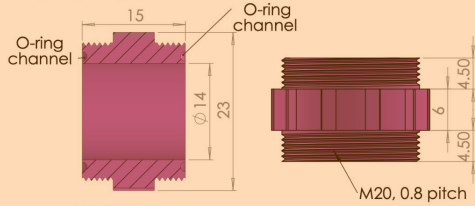
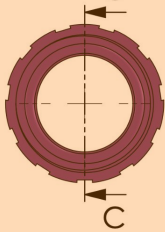
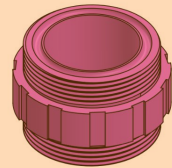
Perspectives - high dose rate (FLASH regime)



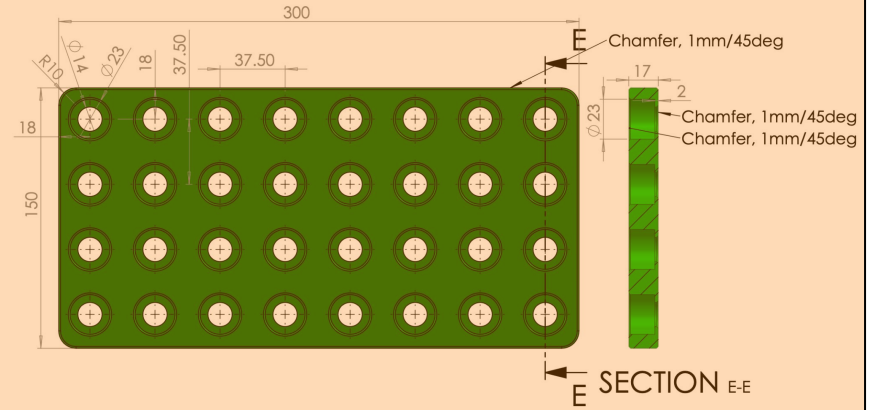
SECTION A-A



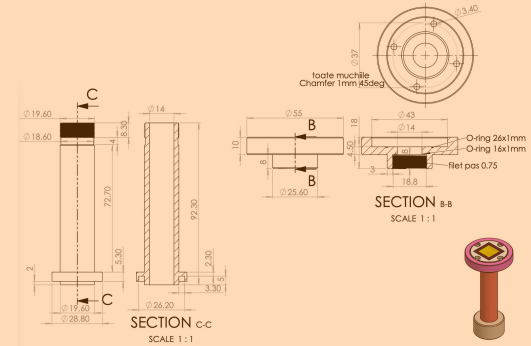
SECTION D-D



SECTION C-C



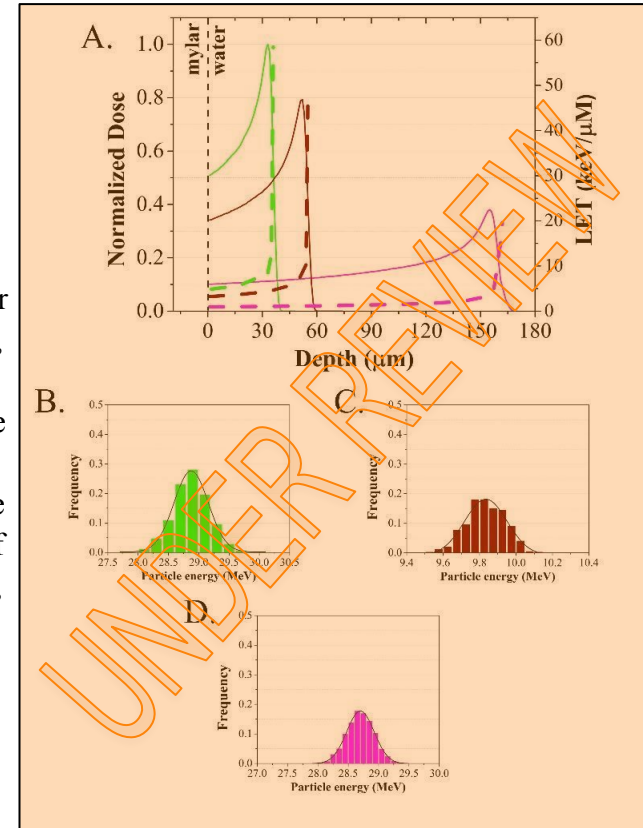
SECTION E-E



Courtesy of A. Enciu



- A. Geant4 simulation of dose (line plot) and LET (dashed plot) in water after heavy ions propagation through the 2 μm Al thin foil; green: $^{12}\text{C}^{6+}$ of 56 MeV, brown: $^7\text{Li}^{3+}$ of 32 MeV, magenta: $^{11}\text{B}^{5+}$ of 48 MeV;
- B. Energy distribution of the 56 MeV $^{12}\text{C}^{6+}$ beam at the mylar-water interface (mean energy = 28.87 MeV, FWHM = 679.63 keV);
- C. Energy distribution of the 48 MeV $^{11}\text{B}^{5+}$ beam at the mylar-water interface (mean energy = 9.83 MeV, FWHM = 245.14 keV);
- D. Energy distribution of the 32 MeV $^7\text{Li}^{3+}$ beam at the mylar-water interface (mean energy = 28.7 MeV, FWHM = 523.62 keV);





Dissemination & References

1. G. Veliş, R. F. Andrei, I. Burducea, A. Enciu, D. Iancu, D. A. Mirea, A. Spiridon, M. Straticiu, **Joint research activities at the 3 MV Tandatron™ from IFIN-HH**, *Eur. Phys. J. Plus* 136 (2021) 1171. <https://doi.org/10.1140/epjp/s13360-021-02156-7>.
2. Mihai Straticiu, Mihaela Bacalum, Calin Mircea Rusu, Radu Andrei, Ion Burducea, Ioan Cenusă, Constantin Cenusă, Irina Dinescu, Simona Dirleci, Alexandru Enciu, Decebal Iancu, Radu Vasilache, Mina Raileanu, Mihai Radu, **New setup for basic radiobiology studies using a 3 MV Tandatron™: design and developments**, NIM B, under review

Nuclear Instruments & Methods in Physics Research, Section B

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Page: 1 of 1 (1 total submissions) Display 10 results per page.

Action	Manuscript Number	Title	Authorship	Initial Date Submitted	Status Date	Current Status
Action Links	V2106	New setup for basic radiobiology studies using a 3 MV Tandatron™: design and developments	Other Author	Sep 14, 2021	Sep 15, 2021	Under Review

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- [1] B. Jones, S. J. McMahon, and K. M. Prise, “The Radiobiology of Proton Therapy: Challenges and Opportunities Around Relative Biological Effectiveness,” *Clin. Oncol. (R. Coll. Radiol.)*, vol. 30, no. 5, pp. 285–292, May 2018, doi: 10.1016/j.clon.2018.01.010.
- [2] H. Paganetti, *Proton Beam Therapy*. IOP Publishing, 2017.
- [3] K. Ilicic, S. E. Combs, and T. E. Schmid, “New insights in the relative radiobiological effectiveness of proton irradiation,” *Radiat. Oncol.*, vol. 13, no. 1, p. 6, 2018, doi: 10.1186/s13014-018-0954-9.
- [4] I. Burducea et al., “A new ion beam facility based on a 3MV Tandatron™ at IFIN-HH, Romania,” *Nucl. Instruments Methods Phys. Res. Sect. B Beam Interact. with Mater. Atoms*, vol. 359, pp. 12–19, 2015, doi: 10.1016/j.nimb.2015.07.011.
- [5] N. Podaru, F. Hoef, A. Gottgang, and D. Mous, “Design and performance of the HVE electrostatic nuclear microprobe,” *Nucl. Instruments Methods Phys. Res. Sect. B Beam Interact. with Mater. Atoms*, vol. 306, pp. 25–28, 2013, doi: 10.1016/j.nimb.2012.11.044.
- [6] R. A. Vasilache, M. A. Popovici, M. Straticiu, M. Radu, and A. Groza, “THE DEVELOPMENT OF A NOVEL ARRAY DETECTOR FOR OVERCOMING THE DOSIMETRY CHALLENGES OF MEASURING IN VERY SHORT PULSED CHARGED PARTICLE BEAMS: THE ELIDOSE PROJECT,” *Radiat. Prot. Dosimetry*, vol. 183, no. 1–2, pp. 285–289, May 2019, doi: 10.1093/rpd/ncy253.
- [7] S. Agostinelli et al., “Geant4—a simulation toolkit,” *Nucl. Instruments Methods Phys. Res. Sect. A Accel. Spectrometers, Detect. Assoc. Equip.*, vol. 506, no. 3, pp. 250–303, 2003, doi: [https://doi.org/10.1016/S0168-9002\(03\)01368-8](https://doi.org/10.1016/S0168-9002(03)01368-8).
- [8] M. Durante, E. Bräuer-Krisch, and M. Hill, “Faster and safer? FLASH ultra-high dose rate in radiotherapy,” *Br. J. Radiol.*, vol. 91, no. 1082, p. 20170628, Feb. 2018, doi: 10.1259/bjr.20170628.

DOSE-RATE MODULATE THE EVOLUTION OF TUMORAL CELLS IN CULTURE AFTER SINGLE DOSE IRRADIATION WITH LOW ENERGY PROTONS

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ABSTRACT

Recent studies underline the great benefits of FLASH technique observed across various animal models, and also report the treatment of subcutaneous T-cell lymphoma in a human patient resulting in complete response and minimal toxicities. However, the details of the mechanism by means of the dose-rate modulate the cells responses remains still unrevealed. We present here preliminary results regarding the effect of dose-rate on some basic cellular processes, when the cells in culture are irradiated by a 1.9 MeV proton beam with doses commonly used in radiotherapy.

MATERIALS AND METHODS

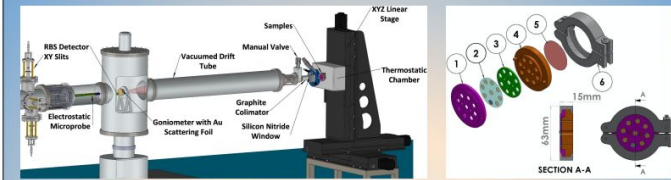


Figure 1 Irradiation facility 3D schematics - left side; drawing of a cell culture chamber (1. Aluminum front cover; 2. Mylar foil; 3. Silicone gasket; 4. Teflon chamber; 5. Breathable sealing tape sterile; 6. KF63 vacuum clamp) - right side

Cell culture

Melanoma cell line, B16 was purchased from ATCC, cultured in Dulbecco's modified Eagle medium (DMEM), and supplemented with 10% fetal bovine serum (FBS) and penicillin-streptomycin (1%-100 units/mL) (Biocrom). This cell line was seeded in special chambers which were manufactured in our institute out of steel, and the cells were grown on a smooth surface made of mylar (polyethylene terephthalate). Cultures were maintained in a humidified atmosphere of 95% air/5% CO₂ at 37°C.

Proton beam was delivered by a 3 MV Tandatron™ located in the Applied Nuclear Physics Department at IFIN-HH [1]. The cells were exposed to 1 and 3 Gy with a dose rate of 5 Gy/s, 50 Gy/s and 250 Gy/s. In order to achieve the FLASH regime a beam electrostatic deflector was used [2].

Beam dosimetry



Figure 2 A. Radiochromic (RCF) film irradiation:

1. SiN₂ window holder; 2. GAFChromic HD-V2 [3] film and 3. PTW Markus ionization chamber [4]; B. Lateral uniformity of the RCF; C. Photo taken under microscope of a Tatrak detector after etching procedure was applied [5]; D. Electrostatic X-sterers (left) and 1 kV power supply triggered by a function generator (right).

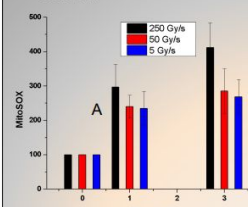
REFERENCES

- [1] I. Burducescu et al., (2015) Nucl. Instrum. Methods Phys. Res. Section B, p. 12-19
- [2] J. D. Wilson et al., (2020) Front Oncol., 9: 1563
- [3] <http://www.gafchromic.com/gafchromic-film/radiotherapy-film/HD-V2/index.asp>
- [4] <https://www.ptw.com/en/products/advanced-markus-electron-chamber/>
- [5] <https://www.tatd.co.uk/index.php>

CONCLUSIONS

1. As expected, the response of the cells to accelerated protons is depending on the dose. All the investigated parameters (ROS production, pre-apoptotic cells level and senescence index) are increasing with the dose.
2. The new information is given by the influence of the dose-rate. At least in our hand, the highest dose-rate investigated here (250 Gy/s) induced the strongest cellular response for all the investigated parameters.

RESULTS



ROS Measurements

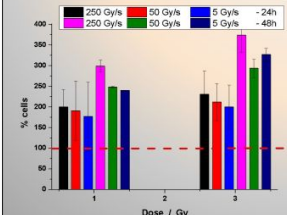
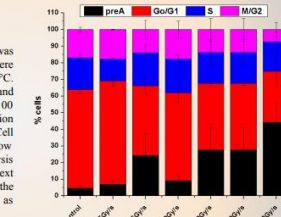
After the irradiation experiments, the cells investigated using MitoSOX (MitoSOX™ Red reagent, Thermo Fisher Scientific, Waltham, Massachusetts, USA). Cells were incubated with 5 μM solution for 30 minutes at 37 °C. This reagent is a fluorescent indicator of superoxide molecule produced by the mitochondria. After the incubation period, the solution is removed and the cells were lysed with trypsin. Finally, the fluorescence of the samples was measured using a plate reader (Mithras LB 940) at λ_{exc} = 485 nm/λ_{em} = 590 nm. Data is represented as mean ±SD (n = 3).

Figure 3: ROS measurements

Cell Cycle Analysis

Cell cycle analysis (B16 cell population distribution) was analysed after 24 h of proton irradiation. The cells were harvested and fixed with pre-chilled 70% ethanol at -20°C. Following, the cells were incubated with 0.2 mg/ml RNase and 20 μg/ml propidium iodide (PI) in a 0.1% Triton TX-100 solution in the dark for 30 min at 37 °C. Cell cycle distribution was analysed by flow cytometry using a Beckman Coulter Cell Lab Quanta SC Flow Cytometer, 771917 Laser, Arc, MPL flow cytometer and data analysed using the Quanta Analysis software. The graph represents cell distribution in the next cellular stages: pre-apoptotic, G0/G1, S and G2/M for the control and the irradiated cells. All the data is represented as mean ±SD (n = 3).

Figure 4: Cell Cycle Analysis



Senescence

After 24 h and 48 h of proton irradiation, the cells were harvested and stained with CellEvent™ Senescence Green according to the protocol. The cells fluorescence was measured using a plate reader (Mithras LB 940) at λ_{exc} = 485 nm/λ_{em} = 590 nm, and the results were normalized to the control cells.

Figure 5: Senescence index in cells irradiated with protons after 24h (1) and 48h (3).

Acknowledgements

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mai ales pentru reziliență!*



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